



How Trichoderma Enhances Plant Defense Enzymes in Food Crops: Current Insights and Future Perspectives in Asia

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Received: 21 October 2024

1st Revised: 08 December 2024

Accepted: 27 December 2024

Published: 30 December 2024

Abstract

Food crops are plants grown for consumption by humans and animals. These crops face threats from both abiotic and biotic factors. Therefore, environmentally friendly alternative controls, such as enhancing plant resistance using *Trichoderma* spp, are needed. This systematic literature review provides information on the effects of *Trichoderma* spp. application on plant physiology, particularly focusing on defense enzymes in food crops in the Asian region. Articles on the effects of *Trichoderma* application on food crop defense enzymes in Asia were retrieved from PubMed and Scopus. The literature search yielded 3,022 articles from these databases. Four criteria were established for study inclusion: i) studies involving *Trichoderma* spp.; ii) studies using food crops as the subject; iii) studies conducted in Asia; and iv) studies evaluating at least one defense enzyme activity. In this systemic review, we found at least 100 isolates of *Trichoderma* spp. that can increase plant resistance, as indicated by the increase of plant defense enzymes in Asia. The studies were mostly reported in India and China. Rice is the most frequently studied crop. The most frequently used is *T. harzianum*. Meanwhile, the defense enzymes tested are peroxidase, polyphenol oxidase, phenylalanine ammonia-lyase, chitinase, and β -1,3-glucanase. The most commonly used inoculation method is seed treatment in a controlled environment such as a greenhouse. More research on changes in plant enzymes due to *Trichoderma* spp. application is recommended to assess its effectiveness in practical, real-world settings, especially in field conditions.

Keywords: Food crops, induce systemic resistance, plant defense enzymes, *Trichoderma* spp.

Introduction

Food crops are essential agricultural commodities grown for human and animal consumption (Aly & Borik, 2023). The growth and production of food crops have become a primary focus in global agriculture due to their crucial role in food security (Guleria et al.,

2023). In 2021, corn and rice ranked as the second and third most-produced agricultural commodities worldwide, following sugarcane (Statista, 2023). However, food crops are continuously threatened by various abiotic factors such as water scarcity, soil pollution, and climate change, as well as biotic factors,

including pest and disease attacks (Guleria et al., 2023).

Nowadays, the most widely used pest and disease management strategies involve resistant crop varieties and chemical pesticides. However, these methods have limitations. Pests and pathogens are highly adaptable, resilient, and capable of evolving resistance to pesticides, especially under changing climatic conditions (Kaur & Chauhan, 2023). Additionally, excessive pesticide use in agriculture poses significant environmental risks, including soil and water contamination and non-target organism toxicity. The rapid increase in pesticide resistance further exacerbates this issue, reducing the efficacy of previously effective pesticides (Kaur & Chauhan, 2023). Therefore, alternative environmentally friendly control strategies are urgently needed, such as enhancing plant resistance through biological agents like *Trichoderma* spp.

Trichoderma spp. are beneficial microorganisms known for their role in improving soil fertility and plant growth. These fungi establish mutualistic endophytic relationships with various plant species, enhancing host plant development, providing protection against pathogen attacks, and improving nutrient uptake efficiency (Awad-Allah et al., 2023). The effectiveness of *Trichoderma* spp. in mitigating abiotic and biotic stress has been widely documented (Zaidi et al., 2014; Gangwar & Singh, 2018). However, most studies evaluating plant resistance induction by *Trichoderma* spp. focus primarily on visual damage assessments caused by abiotic and biotic stresses. Limited research has explored the physiological mechanisms underlying *Trichoderma*-induced resistance, particularly changes in plant defense enzyme activity.

This literature review aimed to examine the impact of *Trichoderma* spp. application on

plant physiology, with a specific focus on the activity of defense enzymes in food crops in Asia. By analyzing the physiological responses of food crops to *Trichoderma* spp., this study seeks to provide deeper insights into the mechanisms of plant resistance induction. The findings are expected to contribute to the development of sustainable agricultural practices and optimize the use of *Trichoderma* spp. as an effective and environmentally friendly biological control agent.

Methods

Database Sources

Articles reviewed on the effect of *Trichoderma* application on food crop defence enzymes in Asia were retrieved from online databases at 21 September 2024. PubMed and Scopus were used as the databases. The search was limited to English-language journal articles with publication dates between 2014 and 2024.

Search Term

The following search strings were utilized in the selection of relevant articles in the title, keywords, or abstract: ("Boost*" OR "Enhanc*" OR "Increas*" OR "Improv*" OR "Stimulat*" OR "Promot*") AND ("Plant* defens* enzyme*" OR "Plant* Enzyme*" OR "Enzyme*") AND ("Trichoderma*") AND ("Agricultural field*" OR "Field*" OR "Farmland*"). The literature search yielded 3022 records from Scopus and Pubmed, which were imported into Rayyan AI (<http://rayyan.qcri.org>) to manage and screen the data.

Screening and Data Extraction

The screening process removed duplicates, conference proceedings, reviews, book chapters, and editorial material. The remaining records were retrieved in full text and reviewed in detail. Screening was carried out against the inclusion/exclusion criteria. Four criteria were set for study inclusion: i)

studies with *Trichoderma* spp.; ii) studies using food crops as an object; iii) studies conducted in Asia; and iv) studies evaluating at least one increasing enzymes defense activity (β -1,3-glucanase, Phenylalanine ammonia-lyase, Peroxidase, Chitinase, Chitosanase, and Polyphenol oxidase). Study exclusion criteria were: i) studies not examine plant defense enzymes; ii) there is no interaction between *Trichoderma* spp. and

food crop plants; iii) studies not written in English; iv) Introduction of *Trichoderma* spp. in combination with other microbes (unable to see the effects of *Trichoderma* directly). This systematic review followed the PRISMA statement and checklist guidelines for screening publications (Moher et al., 2009). A checklist for the systematic review is provided in Figure 1.

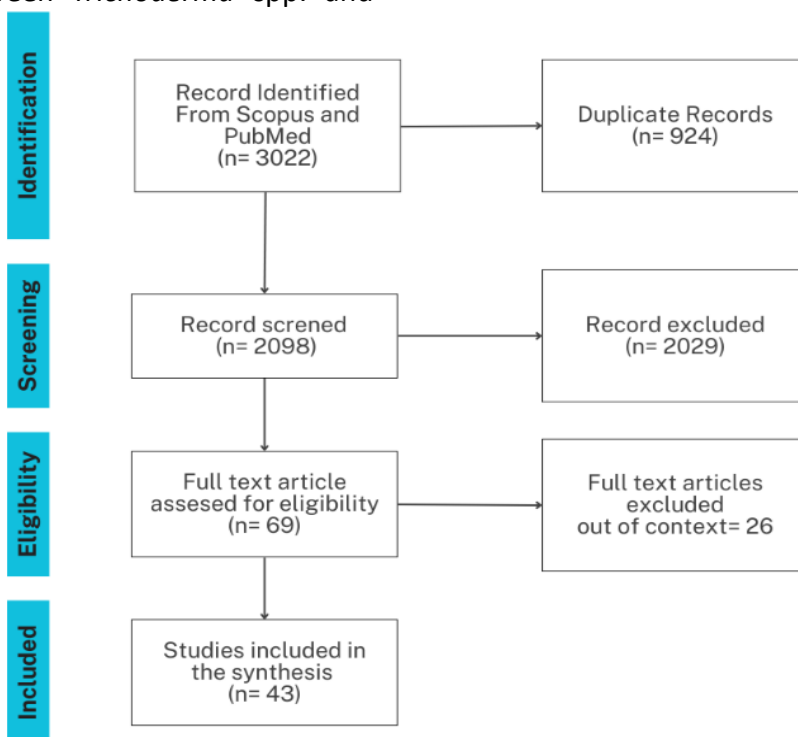


Figure 1. Preferred reporting items for systematic reviews and meta-analysis (PRISMA) diagram of systematic quantitative literature review selection process. Selection criteria are outlined in Section 2.3. represents the number of publications

Potentially relevant articles were downloaded and read in full. Information and data relevant to this review were extracted from eligible articles. The information included the Geographical distribution, the effect of *Trichoderma* sp. on plant defense enzymes, inoculation methods, and the study setting. The study settings used in each article were categorized into controlled and uncontrolled environments (e.g., greenhouse and field). All figures were constructed using Microsoft Excel 2021 and mapchart.net

Results

The geographical distribution of studies measuring the enhancement of defense enzyme activity in food crops using the biocontrol agent *Trichoderma* spp. in Asia reveals that India has conducted the most research on this topic, with a total of 26 studies. This is followed by China, Pakistan, Iraq, Iran, Thailand, and Malaysia (Figure 2). The most common inoculation method is seed treatment (Figure 3)



Figure 2. Geographical distribution of *Trichoderma* spp. application for enhancing food crops defense enzymes in Asia

A total of 100 isolates were identified across 43 reviewed articles. The most frequently used *Trichoderma* species was *T. harzianum*, appearing 43 times. Meanwhile, rice was the most commonly studied crop for

enhanced defense enzyme activity. We also found that the defense enzymes tested were peroxidase, polyphenol oxidase, phenylalanine ammonia-lyase, chitinase, and β -1,3-glucanase (Table 1).

Table 1. *Trichoderma* species effect on plant defense enzyme

<i>Trichoderma</i> species	Plant species	Plant defense enzyme effected	References
<i>T. harzianum</i> T-14	Rice	Increasing 27.07 % POD activity than control	Datta et al., 2022
<i>T. harzianum</i> 94(A)		Increasing 23.29 % POD activity than control	
<i>T. harzianum</i> IRRI-2		Increasing 46.64 % POD activity than control	
<i>T. asperellum</i> OT-3		Increasing 27.30 % POD activity than control	
<i>T. harzianum</i> OT-8		Increasing 27.09 % POD activity than control	
<i>T. harzianum</i>	Chickpea	POD activities were significantly higher than the control	Javaid et al., 2021
<i>T. viride</i>		POD activities were significantly higher than the control	
<i>T. asperellum</i>	Maize	POD activities were significantly higher than the control treatment with meadow soil	Fu et al., 2017

<i>Trichoderma</i> species	Plant species	Plant defense enzyme effected	References
<i>T. viride</i> , <i>T. harzianum</i> , <i>T. harzianum</i> Th14, <i>T. harzianum</i> Th19, <i>T. harzianum</i> ThR, <i>T. asperellum</i> TUK42	Shorgum	POD, PPO, PAL activities were significantly higher than the control	Yadav et al., 2023
<i>T. harzianum</i> <i>T. viride</i>	Cowpea	Increasing activity of POD, PPO, and PAL than the control with 41%, 45%, and 39% respectively Increasing activity of POD, PPO, and PAL than the control with 15%, 30%, and 32% respectively	Shoaib et al., 2018
<i>T. longibrachiatum</i>	Wheat	POD activities were significantly higher than the control	Zhang et al., 2016
<i>T. erinaceum</i> T51 <i>T. viride</i> T52	Bean	Significantly Increase activity of PAL after 24, 48, 96 hours after inoculation than the control	Kumar et al., 2024
<i>T. afroharzianum</i> IIPRTh-33 <i>T. asperellum</i> IIPRTh-31	Pigeonpea	POD, PPO, PAL activities were significantly higher than the control POD, PPO, PAL activities were significantly higher than the control	Mishra et al., 2023
<i>T. harzianum</i> IIPRTh-10		POD and PAL activities were significantly higher than the control	
<i>T. harzianum</i> IIPRTh-23		PAL activities were significantly higher than the control	
<i>T. harzianum</i> IIPRTh-12		PAL activities were significantly higher than the control	
<i>T. asperellum</i> IIPRTas-9		POD and PAL activities were significantly higher than the control	
<i>T. asperellum</i> IIPRTas-15		POD and PAL activities were significantly higher than the control	
<i>T. asperellum</i> IIPRTas-12		PAL activities were significantly higher than the control	
<i>T. harzianum</i> IIPRTh-38		POD and PAL activities were significantly higher than the control	
<i>T. harzianum</i> IIPRTh-22		PAL activities were significantly higher than the control	
<i>T. asperellum</i> IIPRTas-13		POD and PAL activities were significantly higher than the control	
<i>T. harzianum</i> IIPRTh-3		POD and PAL activities were significantly higher than the control	
<i>T. longibrachiatum</i> IIPRTg-3		PAL activities were significantly higher than the control	
<i>T. asperellum</i> IIPRTas-1		POD and PAL activities were significantly higher than the control	
<i>T. harzianum</i> IIPRTh-39		POD and PAL activities were significantly higher than the control	
<i>T. asperellum</i> T4 <i>T. harzianum</i> T6 <i>T. asperellum</i> T19	Great millet	POD and PPO activities were significantly higher than the control POD and PPO activities were significantly higher than the control POD activities were significantly higher than the control	Manzar et al., 2021

<i>Trichoderma</i> species	Plant species	Plant defense enzyme effected	References
<i>T. asperellum</i> T15		PPO activities were significantly higher than the control	
<i>T. harzianum</i> THU0816 ARS	Bean	POD, PPO, PAL activities were significantly higher than the control	Ram et al., 2015
<i>T. erinaceum</i>	Rice	PPO activities were significantly higher than the control	Shasmita et al., 2019
<i>T. harzianum</i> FCBP-SF-1277	Pea	POD activities were significantly higher with all metabolite concentrations (40, 50, 60%) than the control	Rauf et al., 2024
<i>T. longibrachiatum</i>	Wheat	POD, PPO, PAL activities were significantly higher than the control	Zhang et al., 2014
<i>T. viride</i> <i>T. harzianum</i> <i>T. harzianum</i> Th14, <i>T. harzianum</i> Th19, <i>T. harzianum</i> ThR, <i>T. asperellum</i> TUK42	Sorghum	POD, PPO, PAL activities were significantly higher than the control	Yadav et al., 2023
<i>T. hamatum</i> UoM 13	Pearl Millet	POD, PPO, PAL, and β -1,3-glucanase activities were significantly higher than the Untreated Plant	Siddaiah et al., 2017
<i>T. harzianum</i>	Broad bean	PAL activities were significantly higher than the control	Al-Ameri, 2023
<i>T. viride</i> BHU-2953	soybean	<i>T. viride</i> BHU-2953 with addition of recommendation of fertilizer was significantly increasing the activity of POD enzymes than the control	Paul and Rakshit, 2022
<i>T. harzianum</i> IIPRTh-3	Pigeonpea	POD and PAL activities were significantly higher than the control	Mishra et al., 2024
<i>T. aureoviride</i> FCB 691	Mungbean	Soil application with <i>T. aureoviride</i> + biomass of <i>Coronopus didymus</i> and inoculated with <i>Macrophomina phaseolina</i> can increased POD, PPO, and PAL significantly than the control	Javaid et al., 2018
<i>T. viride</i>	Potato	POD activities were significantly higher than the control after 72 and 96 hours after pathogen inoculation PPO activities were significantly higher than the control after 48, 72, and 96 hours after pathogen inoculation PAL activities were significantly higher than the control after 48, 72, and 96 hours after pathogen inoculation	Kumar et al., 2023
<i>T. asperellum</i> T42	Rice	POD and PAL activities were significantly higher than the Untreated Plant in Drought or Non-Drought conditions	Singh et al., 2020
<i>T. paraviridescens</i> <i>T. erinaceum</i> <i>T. asperellum</i>	Rice	PPO activities were significantly higher than the Untreated Plant PPO activities were significantly higher than the Untreated Plant PPO and POD activities were significantly higher than the Untreated Plant	Islam et al., 2023

<i>Trichoderma</i> species	Plant species	Plant defense enzyme effected	References
<i>T. harzianum</i> CRRI-T1 <i>T. erinaceum</i> CRRI-T2 <i>T. atroviride</i> CRRI-T3 <i>T. atroviride</i> CRRI-T5 <i>T. atroviride</i> CRRI-T9 <i>T. atroviride</i> CRRI-T13	rice	PPO activities were significantly higher in shoot and root than the Untreated Plant	Swain et al., 2018
<i>T. harzianum</i> Th9	Potato	Chitinase and β -1,3-glucanase activities were significantly higher than the control	Singh et al., 2020
<i>T. harzianum</i> UPMT1 <i>T. virens</i> UPMT2	Rice	POD, PPO, and PAL activities were significantly higher in shoot and root than the Untreated Plant POD, PPO, and PAL activities were significantly higher in shoot and root than the Untreated Plant	Ng et al., 2015
<i>T. harzianum</i>	Maize	PPO activities were significantly higher than the untreated plant	Afrouz et al., 2023
<i>T. harzianum</i> 56 <i>T. harzianum</i> 69 <i>T. harzianum</i> 75 <i>T. harzianum</i> 82 <i>T. harzianum</i> 89	Wheat	PAL activities were significantly higher than the untreated plant after 13 days treated with drought stress	Shukla et al., 2014
<i>T. harzianum</i> CRRI-T1 <i>T. erinaceum</i> CRRI-T2 <i>T. artroviride</i> CRRI-T3 <i>T. hebeiensis</i> CRRI-T15 <i>T. parareesei</i> CRRI-T16 <i>T. longibrachiatum</i> CRRI-T22 <i>T. reesei</i> CRRI-T27	Rice	POD and PPO activities were significantly higher than the control	Swain et al., 2021
<i>T. harzianum</i> <i>T. viride</i>	Rice	POD activities were significantly higher than the control	Ghosh, 2017
<i>T. asperellum</i> GDFS1009	Maize	POD and PPO activities were significantly higher than the control 0 and 72 hours after inoculation	Batool et al., 2020
<i>T. harzianum</i>	Pea	PAL activities were significantly higher than the control when measured 24 & 48 hours after inoculation PPO activities were significantly higher than the control when measured 96 hours after inoculation POD activities were significantly higher than the control when measured 24 hours after inoculation	Pandey et al., 2021
<i>T. hamatum</i> UoM 13	Pearl Millet	POD, PPO, PAL and β -1,3-glucanase activities were significantly higher than the control	Lavanya et al., 2017
<i>T. harzianum</i>	Wheat	POD activities were significantly higher than the control	Younis et al., 2022
<i>T. artroviride</i>	Pearl Millet	POD activities were significantly higher than the control	Nandini et al., 2017
<i>T. asperellum</i> T42	Pea	PAL activities were significantly higher than the control	Patel et al., 2017

<i>Trichoderma</i> species	Plant species	Plant defense enzyme effected	References
<i>T. asperelloides</i> PSU-P1	Rice	POD activities in shoots and roots plant treated with <i>T. asperelloides</i> were significantly higher than the control PAL activities in roots plant treated with <i>T. asperelloides</i> were significantly higher than the control	Wonglom et al., 2024
<i>T. harzianum</i> UBSTH-501	Wheat	POD and PAL activities were significantly higher than the control	Singh et al., 2019
<i>T. harzianum</i> T891	kidney bean	POD activities were significantly higher than the control	Guo et al., 2024
<i>T. longibrachiatum</i> TG1	Wheat	POD and PAL activities were significantly higher than the control	Boamah et al., 2021
<i>T. longibrachiatum</i> TG1	Wheat	POD and PAL activities were significantly higher than the control	Boamah et al., 2022
<i>Trichoderma</i> CRRIT-5	Rice	POD activities in roots were significantly higher than the control	Sarangi et al., 2021
<i>Trichoderma</i> CRRIT-13		POD activities in shoots and roots were significantly higher than the control	
<i>Trichoderma</i> NRRIT-26		POD activities in shoots and roots were significantly higher than the control	
<i>Trichoderma</i> NRRIT-27		POD activities in shoots and roots were significantly higher than the control	
<i>T. viride</i>	Potato	PAL activities were significantly higher than the control after 0, 24, 48, and 72 hours after pathogen inoculation β -1,3-glucanase activities were significantly higher than the control after 24, 48, and 72 hours after pathogen inoculation	Kumar et al., 2021
<i>T. harzianum</i>		PAL activities were significantly higher than the control after 0, 24, 48, and 72 hours after pathogen inoculation β -1,3-glucanase activities were significantly higher than the control after 0, 24, 48, and 72 hours after pathogen inoculation	
<i>T. reesei</i>	Rice	PPO and PAL activities in shoots and roots were significantly higher than the control	Singh et al., 2019

Note: POD= Peroxidase, PAL= Phenylalanine ammonia lyase, and PPO= Polyphenol oxidase

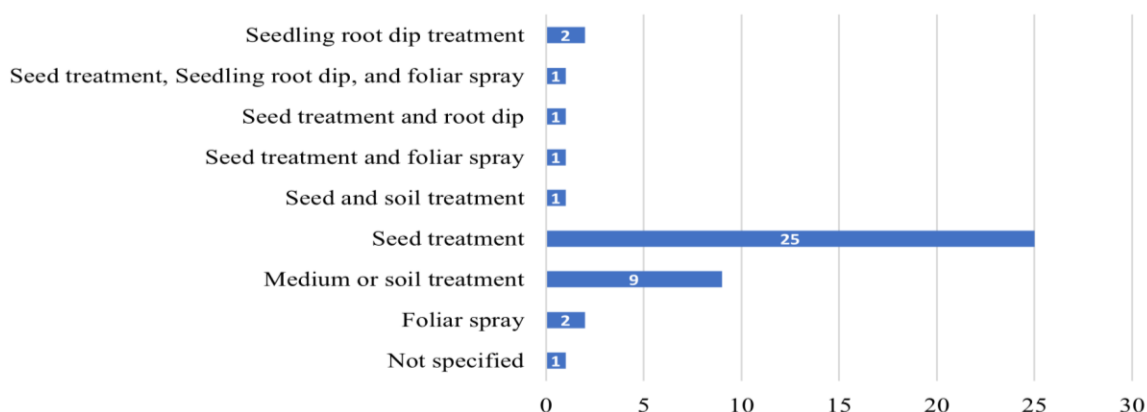


Figure 3. Inoculation methods of *Trichoderma* spp.

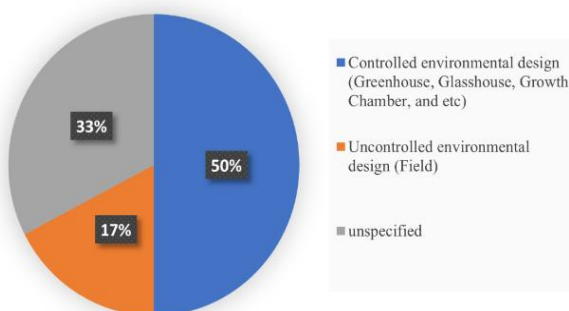


Figure 4. Study setting of *Trichoderma* spp. affecting plant defense enzyme

Discussion

Studies measuring the increase of defense enzyme activity in Asian food crops using *Trichoderma* spp. have been mostly reported in India and China (Figure 2). This is closely related to their superior research capabilities compared to other Asian countries. According to Scimago (2024), data on scientific publications in 2023 show that India and China rank among the top three countries with the highest number of scientific publications worldwide.

Rice is the most frequently studied crop (Table 1). This is linked to the cultural habits of India and China, where rice is a staple food. In 2023–2024, China and India were the largest consumers of rice globally (Statista, 2024). Rice is the most researched commodity and as approximately 90% of global rice production and consumption happens in Asia (Bandumula, 2017).

The most frequently used *Trichoderma* species was *T. harzianum*, appearing 43 times. Meanwhile, defense enzymes tested were peroxidase, polyphenol oxidase, phenylalanine ammonia-lyase, chitinase, and β -1,3-glucanase (Table 1). Plant induced resistance is linked to these defense enzymes. Every plant defence enzyme has its own mechanism for enhancing resistance. For example, the enzyme peroxidase. Pathogen or biocontrol agent infection in host plant tissues stimulates peroxidases, a well-known class of PR

proteins. They are able to limit the cellular spread of infection by forming structural barriers or by creating highly toxic environments by massive ROS and RNS generation (Appu et al., 2021).

There is a strong correlation between increased plant defense enzyme activity due to *Trichoderma* spp. application (Kumar et al., 2023; Gashash et al., 2023). *Trichoderma* spp. increases plant defence enzymes with its ability to be symbiotic with plants. *Trichoderma* spp. acts as an elicitor that can make plants respond to the stimulus given to them, thus this *Trichoderma* treatment enhances plant defence enzymes that will lead to the formation of jasmonic acid, salicylic acid and other defence metabolites such as phenol, lignin, phytoalexins, and others. Therefore, the plants are more resistant if there is infection from pathogens afterwards (Islam et al., 2023) (Yao et al., 2023).

The most common inoculation method is seed treatment (Figure 3). The method plays a vital role in its effectiveness in influencing plant resistance. It also considers factors such as the type of biocontrol agent, the pathogen, and the specific plant species studied (Lavanya et al., 2017). It is regarded as easier compared to other methods like root soaking or inoculation in the growing medium (Kumar et al., 2014). This suggests that treating seeds with *Trichoderma* spp. can enhance plant resistance, evidenced by increased plant defense enzyme activity. Therefore, for future applications of *Trichoderma* sp. it is highly recommended to apply it to seeds.

Most studies have been conducted in controlled environments such as greenhouse (Figure 4). The place where research is conducted also plays a crucial role in the outcome of the research. Research conducted in a controlled environment tends to provide better results than research in an uncontrolled

environment. In addition, data bias is also more common in studies in uncontrolled environments than in controlled environments (Kandi and Vadakedath, 2023). However, research on uncontrolled environments in agriculture must still be conducted by following proper research principles in order to provide many benefits to farmers who generally carry out agricultural practices in the field (Dechassa et al., 2022).

Conclusion

The literature search yielded 3,022 articles from these databases. Four criteria were established for study inclusion: i) studies involving *Trichoderma* spp.; ii) studies using food crops as the subject; iii) studies conducted in Asia; and iv) studies evaluating at least one defense enzyme activity. In this systemic review, we found at least 100 isolates of *Trichoderma* spp. that can increase plant resistance, as indicated by the increase of plant defense enzymes in Asia. The studies were mostly reported in India and China. Rice is the most frequently studied crop. The most frequently used is *T. harzianum*. Meanwhile, the defense enzymes tested are peroxidase, polyphenol oxidase, phenylalanine ammonia-lyase, chitinase, and β -1,3-glucanase. The most commonly used inoculation method is seed treatment in a controlled environment such as a greenhouse. More research on changes in plant enzymes due to *Trichoderma* spp. application is recommended to assess its effectiveness in practical, real-world settings, especially in field conditions.

Acknowledgement

We would like to express our sincere gratitude to Prof. Madya Dr. Nur Ain Izzati Binti Mohd Zainudin, Dr. Wan Mohd Syazwan Bin Wan Solahudin, and Dr. Noraini Binti Abu Bakar for their invaluable guidance and

insights throughout the course of this research.

Declaration

Author contribution

Hasan Albana is the main contributor and corresponding author for this paper. Nurbailis and Darnetty are co-author. All authors read and approved the final paper.

Funding statement

This research did not receive any specific grant from funding agencies in the public, commercial, or not-for-profit sectors.

Competing interest

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

References

- Afrouz M, RZ Sayyed, B Fazeli-Nasab, R Piri, W Almalki, BN Fitriatin. 2023. Seed bio-priming with beneficial *Trichoderma harzianum* alleviates cold stress in maize. PeerJ 11: e15644. DOI: 10.7717/peerj.15644
- Al-Ameri HA. 2023. Estimation of phenyl alanine ammonia-lyase (PAL) and some phytoalexins in *Vicia faba* plants infected with spot disease caused by *Alternaria alternata*. Journal of Applied Natural Science 15(4): 1499-1504. DOI: 10.31018/jans.v15i4.4811
- Aly AA, ZM Borik. 2023. Food crops, growth and productivity as an important focus for sustainable agriculture. In: Plant Small RNA in Food Crops. Academic Press. London. DOI: 10.1016/B978-0-323-91722-3.00006-3
- Appu M, P Ramalingam, A Sathiyarayanan, J Huang. 2021. An overview of plant defense-related enzymes responses to biotic stresses. Plant Gene 27: 100302. DOI: 10.1016/j.plgene.2021.100302
- Awad-Allah EF, AH Shams, AA Helaly, and El Ragheb. 2022. Effective applications of

- Trichoderma* spp. as biofertilizers and biocontrol agents mitigate tomato Fusarium wilt disease. *Agriculture* 12 (11): 1950. DOI: 10.3390/agriculture12111950
- Bandumula N. 2018. Rice production in Asia: Key to global food security. *Proceedings of the National Academy of Sciences, India Section B: Biological Sciences* 1323-1328. DOI: 10.1007/s40011-017-0867-7
- Batool R, MJ Umer, Y Wang, K He, T Zhang, S Bai, Z Wang. 2020. Synergistic effect of *Beauveria bassiana* and *Trichoderma asperellum* to induce maize (*Zea mays* L.) defense against the Asian corn borer, *Ostrinia furnacalis* (Lepidoptera, Crambidae) and larval immune response. *International Journal of Molecular Sciences* 21(21): 8215. DOI: 10.3390/ijms21218215
- Boamah S, S Zhang, B Xu, T Li, A Calderón-Urrea. 2021. *Trichoderma longibrachiatum* (TG1) enhances wheat seedlings tolerance to salt stress and resistance to *Fusarium pseudograminearum*. *Frontiers in Plant Science* 12: 741231. DOI: 10.3389/fpls.2021.741231
- Boamah S, S Zhang, B Xu, T Li, A Calderón-Urrea, RJ Tiika. 2022. *Trichoderma longibrachiatum* TG1 increases endogenous salicylic acid content and antioxidants activity in wheat seedlings under salinity stress. *PeerJ* 10: e12923. DOI: 10.7717/peerj.12923
- Datta D, AK Senapati, L Behera, NW Zaidi, S Kumar, P Dey. 2023. Alleviating drought stress in rice plant through intervention of *Trichoderma* spp. *Journal of Environmental Biology* 44(3): 373-379. DOI: 10.22438/jeb/44/3/MRN-4084
- Dechassa N, N Keneni, S Bediye, G Alemayehu. 2022. Research methods: A Handbook for Agricultural Researchers. Ethiopian Institute of Agricultural Research. Addis Ababa. Ethiopia.
- Eman AA, IA Mohamed, SF Allah, AH Shams, IH Elsokkary. 2023. *Trichoderma* species: An overview of current status and potential applications for sustainable agriculture. *Indian Journal of Agricultural Research* 57(3): 273-282. DOI: 10.18805/IJARE.AF-751
- Fu J, Z Liu, Z Li, Y Wang, K Yang. 2017. Alleviation of the effects of saline-alkaline stress on maize seedlings by regulation of active oxygen metabolism by *Trichoderma asperellum*. *PLoS One* 12(6): e0179617. DOI: 10.1371/journal.pone.0179617
- Gangwar OP and AP Singh. 2018. *Trichoderma* as an efficacious bioagent for combating biotic and abiotic stresses of wheat—A review. *Agricultural review* 39(1): 48-54. DOI: 10.18805/ag.R-1770
- Gashash EA, SA El-Sayed, WM Kamel, EA Sarhan. 2023. Evaluation of the potential of native *Trichoderma* spp. against root-rot diseases of soybean. *Archives of Phytopathology and Plant Protection* 56(16): 1282-1299. DOI: 10.1080/03235408.2023.2279943
- Ghosh SK. 2017. Study of effects of antagonistic soil microbes on rice early stand establishment, prevention of seedling mortality and induction of peroxidase and catalase enzymes. In: *Plant Cell Biotechnology and Molecular Biology*. Society for Biology and Biotechnology. India
- Guleria P, V Kumar. 2023. *Plant small RNA in food crops*. Elsevier. London. DOI: 10.1016/B978-0-323-91722-3.00006-3
- Guo Z, J Zhang, Z Liu, Y Li, M Li, Q Meng, M Yan. 2024. *Trichoderma harzianum* prevents red kidney bean root rot by increasing plant antioxidant enzyme activity and regulating the rhizosphere microbial community. *Frontiers in Microbiology* 15: 1348680. DOI: 10.3389/fmicb.2024.1348680
- Islam MR, R Chowdhury, AS Roy, MN Islam, MM Mita, S Bashar, MA Latif. 2023. Native *Trichoderma* induced the defense-related enzymes and genes in rice against *Xanthomonas oryzae* pv. *oryzae* (Xoo). *Plants* 12(9): 1864. DOI: 10.3390/plants12091864
- Javaid A, A Ali, A Shoaib, and IH Khan. 2021. Alleviating stress of *Sclerotium rolfsii* on growth of chickpea var. Bhakkar-2011 by *Trichoderma harzianum* and *T. viride*. *JAPS: Journal of Animal and Plant Sciences* 31(6): 119

- 1755-1761. DOI: 10.36899/JAPS.2021.6.0378
- Javaid A, IH Khan, A Shoaib. 2018. Management of charcoal rot of mungbean by two *Trichoderma* species and dry biomass of *Coronopus didymus*. *Planta Daninha* 36: e018182795. DOI: 10.1590/s0100-83582018360100124
- Kandi V, S Vadakedath. 2023. Clinical trials and clinical research: a comprehensive review. *Cureus* 15(2): e35077 DOI: 10.7759/cureus.35077
- Kaur S, BS Chauhan. 2023. Challenges and opportunities to sustainable crop production. In: P Guleria and V Kumar (eds). *Plant Small RNA in Food Crops*. Elsevier. DOI: 10.1016/B978-0-323-91722-3.00006-3
- Kumar S, R Chandra, L Behera, C Keswani, E Sansinenea. 2022. Dual *Trichoderma* consortium mediated elevation of systemic defense response against early blight in potato. *European Journal of Plant Pathology* 1-16. DOI: 10.1007/s10658-021-02431-4
- Kumar S, R Chandra, L Behera, I Sudhir, M Meena, S Singh, C Keswani. 2023. Microbial consortium mediated acceleration of the defense response in potato against *Alternaria solani* through prodigious inflation in phenylpropanoid derivatives and redox homeostasis. *Heliyon* 9(11): e22148. DOI: 10.1016/j.heliyon.2023.e22148
- Kumar S, V Shukla, YN Tripathi, M Aamir, K Divyanshu, M Yadav, RS Upadhyay. 2024. Biochemical changes, antioxidative profile, and efficacy of the bio-stimulant in plant defense response against *Sclerotinia sclerotiorum* in common bean (*Phaseolus vulgaris* L.). *Heliyon* 10(1): e23030. DOI: 10.1016/j.heliyon.2023.e23030
- Kumar S, M Thakur, A Rani. 2014. *Trichoderma*: Mass production, formulation, quality control, delivery and its scope in commercialization in India for the management of plant diseases. *African Journal of Agricultural Research* 9(53): 3838-3852. DOI: 10.5897/AJAR2014.%209061
- Lavanya SN, S Niranjana-Raj, SC Nayaka, KN Amruthesh. 2017. Systemic protection against pearl millet downy mildew disease induced by cell wall glucan elicitors from *Trichoderma hamatum* UOM 13. *Journal of Plant Protection Research* 57(3): 1-13. DOI: 10.1515/jppr-2017-0042
- Manzar N, Y Singh, AS Kashyap, PK Sahu, MVS Rajawat, A Bhowmik, AK Saxena. 2021. Biocontrol potential of native *Trichoderma* spp. against anthracnose of great millet (*Sorghum bicolor* L.) from Tarai and hill regions of India. *Biological Control* 152: 104474. DOI: 10.1016/j.biocontrol.2020.104474
- Mishra RK, S Pandey, KK Hazra, M Mishra, SS Naik, A Bohra, and NP Singh. 2023. Biocontrol efficacy and induced defense mechanisms of indigenous *Trichoderma* strains against Fusarium wilt [*F. udum* (Butler)] in pigeonpea. *Physiological and Molecular Plant Pathology* 127: 102122. DOI: 10.1016/j.pmpp.2023.102122
- Mishra RK, S Pandey, M Mishra, U Singh Rathore, B Singh. 2024. Identification and characterization of high salinity tolerant *Trichoderma harzianum* strain, IIPRTh-3 for mitigating salt stress, wilt incidence and promoting plant growth in pigeonpea. *Archives of Phytopathology and Plant Protection* 57(9): 691-704. DOI: 10.1080/03235408.2024.2390212
- Moher D, A Liberati, J Tetzlaff, DG Altman. 2009. Preferred reporting items for systematic reviews and meta-analyses: The Prisma statement. *PLoS Medicine* 6(7): e1000097. DOI: 10.1371/journal.pmed.1000097
- Nandini B, P Hariprasad, HN Shankara, HS Prakash, N Geetha. 2017. Total crude protein extract of *Trichoderma* spp. induces systemic resistance in pearl millet against the downy mildew pathogen. *3 Biotech* 7: 1-10. DOI: 10.1007/s13205-017-0816-4
- Ng LC, M Sariah, O Sariam, O Radziah, MA Zainal Abidin. 2016. PGPM-induced defense-related enzymes in aerobic rice against rice leaf blast caused by *Pyricularia oryzae*.

- European Journal of Plant Pathology 145: 167-175. DOI: 10.1007/s10658-015-0826-1
- Pandey P, KP Kushwaha, V Upadhyay, J Purohit. 2024. Synergistic effect of plant defense elicitors and biocontrol agents on induction of defense enzymes in pea against downy mildew. *Indian Journal of Agricultural Research* 58(1):1-14. DOI: 10.18805/IJARE.A-5798
- Patel JS, RN Kharwar, HB Singh, RS Upadhyay, BK Sarma. 2017. *Trichoderma asperellum* (T42) and *Pseudomonas fluorescens* (OKC)-enhances resistance of pea against *Erysiphe pisi* through enhanced ROS generation and lignifications. *Frontiers in Microbiology* 8: 306. DOI: 10.3389/fmicb.2017.00306
- Paul S, A Rakshit. 2022. Evaluating lignification, antioxidative defense, and physiochemical changes in soybean through bio-priming under graded soil fertilization. *Journal of Soil Science and Plant Nutrition* 22(2): 2295-2306. DOI: 10.1007/s42729-022-00809-9
- Ram RM, AJ Akansha Jain, AS Akanksha Singh, HB Singh. 2015. Biological management of Sclerotinia rot of bean through enhanced host defense responses triggered by *Pseudomonas* and *Trichoderma* species. *Journal of Pure and Applied Microbiology* 9(1):523-532. <https://microbiologyjournal.org/>
- Rauf A, MN Subhani, M Siddique, H Shahid, MB Chattha, AF Alrefaei, RS Lucas. 2024. Cultivating a greener future: Exploiting *Trichoderma* derived secondary metabolites for fusarium wilt management in peas. *Heliyon* 10(7): e29031. DOI: 10.1016/j.heliyon.2024.e29031
- Sarangi S, H Swain, T Adak, P Bhattacharyya, AK Mukherjee, G Kumar, and ST Mehetre. 2021. *Trichoderma*-mediated rice straw compost promotes plant growth and imparts stress tolerance. *Environmental Science and Pollution Research* 28: 44014-44027. DOI: 10.1007/s11356-021-13701-3.
- Scimagojr, 2024. Scimago Journal and Country Rank. <https://www.scimagojr.com/countryrank.php?year=2023>
- Shasmita, H Swain, SK Naik, AK Mukherjee. 2019. Comparative analysis of different biotic and abiotic agents for growth promotion in rice (*Oryza sativa* L.) and their effect on induction of resistance against *Rhizoctonia solani*: a soil borne pathogen. *Biological Control* 133: 123-133. DOI: 10.1016/j.biocontrol.2019.02.013
- Shoaib A, M Munir, A Javaid, ZA Awan, M Rafiq. 2018. Anti-mycotic potential of *Trichoderma* spp. and leaf biomass of *Azadirachta indica* against the charcoal rot pathogen, *Macrophomina phaseolina* (Tassi) Goid in cowpea. *Egyptian Journal of Biological Pest Control* 28: 1-7. DOI: 10.1186/s41938-018-0031-6
- Shukla N, RP Awasthi, L Rawat, J Kumar. 2015. Seed biopriming with drought tolerant isolates of *Trichoderma harzianum* promotes growth and drought tolerance in *Triticum aestivum*. *Annals of Applied Biology* 166(2): 171-182. DOI: 10.1111/aab.12160
- Siddaiah CN, NR Satyanarayana, V Mudili, VK Gupta, S Gurunathan, S Rangappa, RK Srivastava. 2017. Elicitation of resistance and associated defense responses in *Trichoderma hamatum* induced protection against pearl millet downy mildew pathogen. *Scientific Reports* 7(1): 43991. DOI: 10.1038/srep43991
- Singh DP, V Singh, VK Gupta, R Shukla, R Prabha, BK Sarma, JS Patel. 2020. Microbial inoculation in rice regulates antioxidative reactions and defense-related genes to mitigate drought stress. *Scientific Reports* 10(1): 4818. DOI: <https://www.nature.com/articles/s41598-020-61140-w>
- Singh N, A Singh, P Dahiya. 2020. Performance of potent isolate of *Trichoderma harzianum* (Th9) possessing mycoparasitic property against late blight-causing fungus *Phytophthora infestans* in addition to enhancing defensive enzymes activities in potato crop. *Indian Journal of Agricultural Biochemistry* 33(1): 99-101. DOI: 10.5958/0974-4479.2020.00016.7
- Singh SP, S Pandey, N Mishra, VP Giri, S Mahfooz, A Bhattacharya, A Mishra. 2019. Supplementation of *Trichoderma* improves

- the alteration of nutrient allocation and transporter gene expression in rice under nutrient deficiencies. *Plant Physiology and Biochemistry* 143: 351-363. DOI: 10.1016/j.plaphy.2019.09.015
- Singh UB, D Malviya, S Singh, M Kumar, PK Sahu, HV Singh, AK Saxena. 2019. *Trichoderma harzianum* and methyl jasmonate-induced resistance to *Bipolaris sorokiniana* through enhanced phenylpropanoid activities in bread wheat (*Triticum aestivum* L.). *Frontiers in Microbiology* 10: 1697. DOI: 10.3389/fmicb.2019.01697
- Statista, 2024. Rice consumption worldwide in 2023/2024, by country (in 1,000 metric tons). <https://www.statista.com/statistics/255971/top-countries-based-on-rice-consumption-2012-2013/>
- Statista. 2023. Production volume of the most produced food commodities worldwide in 2021, by product. <https://www.statista.com/statistics/1003455/most-produced-crops-and-livestock-products-worldwide/>
- Swain H, T Adak, AK Mukherjee, PK Mukherjee, P Bhattacharyya, S Behera, M Jena. 2018. Novel *Trichoderma* strains isolated from tree barks as potential biocontrol agents and biofertilizers for direct seeded rice. *Microbiological Research* 214: 83-90. DOI: 10.1016/j.micres.2018.05.015
- Swain H, T Adak, AK Mukherjee, S Sarangi, P Samal, A Khandual, NW Zaidi. 2021. Seed biopriming with *Trichoderma* strains isolated from tree bark improves plant growth, antioxidative defense system in rice and enhances straw degradation capacity. *Frontiers in Microbiology* 12: 633881. DOI: 10.3389/fmicb.2021.633881
- Wonglom P, OU Ruangwong, W Poncheewin, S Arikkit, K Rianguwong, A Sunpapao. 2024. *Trichoderma*-bioenriched vermicompost induces defense response and promotes plant growth in Thai rice variety “Chor Khing”. *Journal of Fungi* 10(8): 582. DOI: 10.3390/jof10080582
- Yadav SS, A Arya, V Singh, Y Singh. 2023. Elicitation of native bioprotective microbial agents associated systemic defense responses and plant growth promotion against bacterial stalk rot pathogen in sorghum (*Sorghum bicolor*). *Phytopathology Research* 5(1): 47. DOI: 10.1186/s42483-023-00202-z
- Yadav SS, A Arya, G Tondon, Y Singh. 2023. Antagonistic microbes mediated modulation of systemic resistance and defense related responses against *Dickeya dadantii* causing bacterial stalk rot in *Sorghum bicolor*. *European Journal of Plant Pathology* 167(4): 833-851. DOI: 10.1007/s10658-023-02769-x
- Yao X, H Guo, K Zhang, M Zhao, J Ruan, J Chen. 2023. *Trichoderma* and its role in biological control of plant fungal and nematode disease. *Frontiers in microbiology* 14: 1160551. DOI: 10.3389/fmicb.2023.1160551
- Younis HS, RM Abdullah, SA Hasan, AA Abdul-Sattar. 2022. Systemic resistance indicators study and seed gall nematode disease caused by *Anguina tritici* affecting of biological and varietal treatments on bread wheat (*Triticum aestivum* L.). *International Journal of Agricultural Statistical Science* 18(1): 289-296. DOI:
- Zaidi NW, MH Dar, S Singh, US Singh. 2014. *Trichoderma* species as abiotic stress relievers in plants. in: Mukherjee PK, Babu S, Singh US, and Das SK (eds). *Biotechnology and Biology of Trichoderma*. Elsevier. Amsterdam.
- Zhang S, Y Gan, B Xu. 2014. Efficacy of *Trichoderma longibrachiatum* in the control of *Heterodera avenae*. *BioControl* 59: 319-331. DOI: 10.1007/s10526-014-9566-y.
- Zhang S, Y Gan, B Xu. 2016. Application of plant-growth-promoting fungi *Trichoderma longibrachiatum* T6 enhances tolerance of wheat to salt stress through improvement of antioxidative defense system and gene expression. *Frontiers in Plant Science* 7: 1405. DOI: 10.3389/fpls.2016.01405

How to cite: H Albana, Nurbailis, and Darnetty. 2024. How *Trichoderma* enhances plant defense enzymes in food crops: Current insights and future perspectives in Asia. *JPT: Jurnal Proteksi Tanaman* (Journal of Plant Protection) 8(2): 109 – 122.